

Intricarene, an Unprecedented Trispiropentacyclic Diterpene from the Caribbean Sea Plume *Pseudopterogorgia kallos*

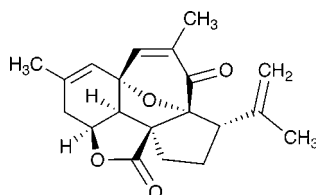
Jeffrey Marrero,[†] Abimael D. Rodríguez,^{*,†} and Charles L. Barnes[‡]

Department of Chemistry, University of Puerto Rico, P.O. Box 23346,
U.P.R. Station, San Juan, Puerto Rico 00931-3346, Department of Chemistry,
125 Chemistry Building, University of Missouri–Columbia, Columbia, Missouri 65211

arodrig@cnnnet.upr.edu

Received March 18, 2005

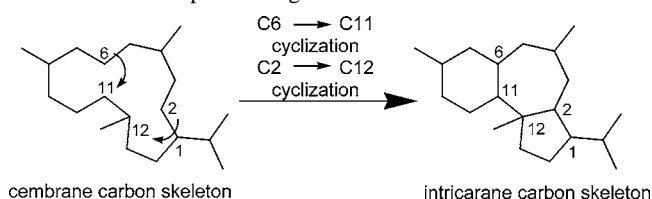
ABSTRACT



A novel trispiropentacyclic diterpene, intricarene (**1**), was isolated from the hexane extract of the Caribbean gorgonian octocoral *Pseudopterogorgia kallos*. Its highly entangled structure was established by interpretation of NMR, IR, UV, and HREIMS data and subsequently confirmed by X-ray diffraction analysis. The unprecedented carbon skeleton of **1** constitutes a new addition to the already impressive architectural diversity of the diterpene class of marine secondary metabolites.

Gorgonian corals of the genus *Pseudopterogorgia* have proven to be a rich source of terpenoid secondary metabolites that frequently exhibit interesting biological activities.¹ As part of an ongoing program to screen marine invertebrate extracts for antitubercular metabolites with low cytotoxicity, we have investigated the MeOH/CHCl₃ extract of a Colombian specimen of the gorgonian coral *Pseudopterogorgia kallos* (Bielschowsky, 1918).² Fractionation of the crude extract led subsequently to the isolation of the novel diterpenoid metabolite, intricarene (**1**), whose complex structure is based on the unprecedented intricarane carbon skeleton (Scheme 1).³ In this paper, we present details of

Scheme 1. Proposed Biogenesis for the Intricarane Skeleton



the isolation, structure determination, proposed biogenesis, and biological activity of **1**.

Specimens of *P. kallos* were collected by hand using scuba on reefs off the coast of Providencia (Old Providence) Island, Colombia, partially air-dried on site, frozen, lyophilized (1.07 kg), cut in small fragments, and homogenized exhaustively with a 1:1 mixture of CH₂Cl₂–MeOH. The green gum obtained (166 g) by evaporating the combined CH₂Cl₂–MeOH extracts was partitioned between hexane and H₂O. The hexane extract was concentrated under reduced pressure,

[†] University of Puerto Rico.

[‡] University of Missouri.

(1) (a) Fenical, W. *J. Nat. Prod.* **1987**, *50*, 1001–1008. (b) Rodríguez, A. D. *Tetrahedron* **1995**, *51*, 4571–4618.

(2) For the first report on the natural products chemistry of *P. kallos*, see: Look, S. A.; Burch, M. T.; Fenical, W.; Qi-tai, Z.; Clardy, J. *J. Org. Chem.* **1985**, *50*, 5741–5746.

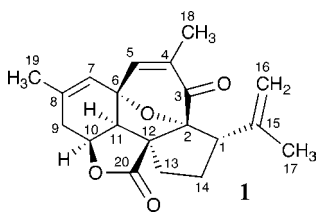
(3) From the Latin word *intricare*, to entangle, perplex.

Table 1. ^1H NMR (500 MHz), ^{13}C NMR (125 Mz), ^1H – ^1H COSY, NOESY, and HMBC Spectral Data for Intricarene (**1**)^a

position	δ_{H} , mult, intrgt (J in Hz)	δ_{C} (mult) ^b	^1H – ^1H COSY	NOESY	HMBC ^c
1	3.38, dd, 1H (11.9, 5.7)	46.4 (CH)	H14 $\alpha\beta$	H13 β , H14 β , H16 β , H ₃ -17	H13 $\alpha\beta$, H16 $\alpha\beta$, H ₃ -17
2		102.8 (C)			H1, H13 α , H14 $\alpha\beta$
3		193.0 (C)			H5, H ₃ -18
4		137.2 (C)			H ₃ -18
5	6.27, q, 1H (1.6)	147.3 (CH)	H ₃ -18	H9 β , H ₃ -18	H11, H ₃ -18
6		84.6 (C)			H5, H10, H11
7	6.41, q, 1H (1.6)	128.1 (CH)	H ₃ -19	H ₃ -19	H9 α , H11, H ₃ -19
8		135.2 (C)			H9 $\alpha\beta$, H ₃ -19
9 α	2.82, dd, 1H (18.6, 8.6)	35.7 (CH ₂)	H9 β , H10	H9 β , H10, H ₃ -19	H7, H11, H ₃ -19
9 β	2.41, br d, 1H (18.6)		H9 α , H10	H5, H9 α	
10	4.77, ddd, 1H (8.6, 5.3, 2.6)	70.7 (CH)	H9 $\alpha\beta$, H11	H9 α , H11	H9 $\alpha\beta$, H11
11	2.54, d, 1H (5.3)	58.0 (CH)	H10	H10, H13 α	H9 $\alpha\beta$, H13 $\alpha\beta$
12		64.2 (C)			H11, H13 β , H14 $\alpha\beta$
13 α	2.01, m, 1H	29.3 (CH ₂)	H13 β , H14 $\alpha\beta$	H11, H13 β	H11
13 β	1.92, m, 1H		H13 α , H14 $\alpha\beta$	H1, H13 α	
14 α	2.11, m, 1H	28.5 (CH ₂)	H1, H13 $\alpha\beta$	H11	H1, H13 $\alpha\beta$
14 β	1.91, m, 1H		H1, H13 $\alpha\beta$	H1	
15		141.8 (C)			H1, H16 β , H ₃ -17
16 α	4.92, q, 1H (1.3)	113.3 (CH ₂)	H16 β , H ₃ -17	H1, H ₃ -17	H ₃ -17
16 β	4.86, br s, 1H		H16 α		
17	1.75, br s, 3H	23.1 (CH ₃)	H16 α	H1, H16 β	H16 $\alpha\beta$
18	1.76, d, 3H (1.6)	14.4 (CH ₃)	H5	H5	H5
19	1.84, t, 3H (1.2)	23.0 (CH ₃)	H7	H7, H9 α	H7, H9 α
20		177.9 (C)			H11, H13 β

^a Spectra were recorded in CDCl_3 at 25 °C. Chemical shift values are in parts per million relative to TMS. ^b ^{13}C NMR multiplicities were obtained from a DEPT-135 experiment. ^c Protons correlated to carbon resonances in ^{13}C column.

and the oily residue (71.9 g) obtained was fractionated via size-exclusion chromatography over Bio-Beads SX-3 in toluene followed by silica gel chromatography using step gradient elution (hexanes to EtOAc) to afford pure intricarene (**1**) as a white crystalline solid (4.0 mg; 0.0024% based on the crude extract dry wt).⁴



Intricarene (**1**) was isolated as optically active crystals ($[\alpha]_{\text{D}}^{20} +50.0^\circ$ (c 0.7, CHCl_3)) that gave an $[\text{M}]^+$ ion at m/z 326.1524 in the HREI-MS appropriate for a molecular formula of $\text{C}_{20}\text{H}_{22}\text{O}_4$ (calcd 326.1518), requiring 10 sites of unsaturation.⁵ The infrared spectrum of **1** showed bands characteristic of γ -lactone and α,β -unsaturated ketone carbonyl groups ($\nu_{\text{max}} = 1767$ and 1690 cm^{-1}). Twenty

resonances ($8 \times \text{C}$; $5 \times \text{CH}$; $4 \times \text{CH}_2$; $3 \times \text{CH}_3$) were observed in the ^{13}C NMR spectrum of **1** (Table 1). Four deshielded resonances in the ^1H NMR spectrum [δ 6.41, q, $J = 1.6\text{ Hz}$ (H-7); 6.27, q, $J = 1.6\text{ Hz}$ (H-5); 4.92, q, $J = 1.3\text{ Hz}$ (H-16 α); 4.86, br s (H-16 β)], three sharp methyl signals [δ 1.84, t, $J = 1.2\text{ Hz}$ (H₃-19); 1.76, d, $J = 1.6\text{ Hz}$ (H₃-18); 1.75, br s (H₃-17)], and six deshielded resonances in the ^{13}C NMR spectrum [δ 147.3 (CH, C-5); 137.2 (C, C-4); 135.2 (C, C-8); 128.1 (CH, C-7); 141.8 (C, C-15); 113.3 (CH₂, C-16)] were readily assigned to three methyl-bearing olefins (two trisubstituted and one 1,1-disubstituted) by analysis of their ^1H scalar coupling patterns and the corresponding ^1H – ^1H COSY, HMQC, and HMBC correlations (Table 1). Two additional carbonyl resonances [δ 193.0 (C-3); 177.9 (C-20)] in the ^{13}C NMR spectrum accounted for five sites of unsaturation, and the absence of ^{13}C NMR evidence for further unsaturated functionality indicated the presence of five additional rings in **1**.

Comprehensive analysis of two-dimensional NMR data, including the results of ^1H – ^1H COSY, HMQC, and HMBC experiments, enabled the complete planar structure of intricarene to be constructed, as in **1**, on a [6–7–5] tricyclic framework substituted with methyl groups and an isopropenyl side chain. To establish these structural features of **1**,

(4) For our prior work in this series, see: (a) Marrero, J.; Rodríguez, A. D.; Baran, P.; Raptis, R. G. *J. Org. Chem.* **2003**, *68*, 4977–4979. (b) Marrero, J.; Rodríguez, A. D.; Baran, P.; Raptis, R. G. *Org. Lett.* **2003**, *5*, 2551–2554. (c) Marrero, J.; Rodríguez, A. D.; Baran, P.; Raptis, R. G.; Sánchez, J. A.; Ortega-Barria, E.; Capson, T. L. *Org. Lett.* **2004**, *6*, 1661–1664. (d) Marrero, J.; Rodríguez, A. D.; Baran, P.; Raptis, R. G. *Eur. J. Org. Chem.* **2004**, 3909–3912.

(5) Data for intricarene (**1**): white crystals; $[\alpha]_{\text{D}}^{20} +50.0^\circ$ (c 0.7, CHCl_3); UV (MeOH) λ_{max} 245 nm (6200); IR (film, NaCl) ν_{max} 3078, 2947, 1767, 1690, 1641, 1627, 1440, 1380, 1282, 1182, 1166, 1002, 973 cm^{-1} ; LREIMS m/z (%) 326 (M^+ , 74), 311 (23), 298 (13), 281 (6), 230 (100), 216 (40), 215 (46), 91 (33); HREI-MS m/z 326.1524, calcd for $\text{C}_{20}\text{H}_{22}\text{O}_4$ 326.1518.

exhaustive NMR measurements were conducted in CDCl_3 . In this solvent, the three methyl signals (a broad singlet for C-17, a doublet for C-18, and a triplet for C-19) were readily resolved. HMBC experiments conclusively placed the methyl groups at strategic locations on the tricarbocyclic ring structure of intricarene. Correlations from the H_3 -17 protons to C-1, C-15, and C-16 placed the isopropenyl side chain at C-1. Correlations from the H_3 -18 methyl protons to C-3 and C-5 allowed this methyl group to be placed at C-4, and likewise, correlations from the H_3 -19 methyl protons to C-7, C-8, and C-9 allowed this methyl group to be placed at C-8. HMBC correlations observed between a carbon resonance at δ 102.8 (C-2) and both the methine resonance at δ 3.38 (H-1) and the methylene resonances at δ 2.01 (H-13 α) and 2.11/1.91 (H-14 $\alpha\beta$) showed that the tertiary carbon C-2, bearing an ether oxygen and a ketone carbonyl, had to be bridged by a saturated ethylene group through C-1 and C-12 to give a cyclopentane ring (C-1 to C-2 and C-12 to C-14). HMBC correlations between the pivotal methine resonance at δ 2.54 (H-11) and all of the ^{13}C resonances at δ 177.9 (C-20), 147.3 (C-5), 128.1 (C-7), 84.6 (C-6), 70.7 (C-10), 64.2 (C-12), 35.7 (C-9), and 29.3 (C-13) demonstrated that such a key bridgehead carbon (C-11) had to be flanked by the spirocyclic carbons C-6 and C-12 and by the oxymethine carbon C-10. A series of HMBC correlations between the olefinic resonance at δ 6.27 (H-5) and the ^{13}C resonances at δ 193.0 (C-3), 84.6 (C-6), and 14.4 (C-18) showed that the spirocyclic carbon atoms C-2 and C-6 had to be bridged by an ether oxygen and the α,β -unsaturated ketone bearing the methyl (C-18) group to give an 8-oxabicyclo[3.2.1]octane ring system (C-2 to C-6 and C-11 to C-12). Similarly, HMBC correlations between δ 6.41 (H-7) and both 35.7 (C-9) and 23.0 (C-19) and between 1.84 (H₃-19) and 135.2 (C-8), 128.1 (C-7), and 35.7 (C-9) accounted for the cyclohexene ring in **1** (C-6 to C-11). Last, HMBC correlations between δ 177.9 (C-20) and both δ 2.54 (H-11) and 1.92 (H-13 β) indicated that C-20 had to be bonded to a quaternary carbon (C-12) and to C-11 through the oxymethine carbon at δ 70.7 (C-10) to complete the final ring required in **1**.

Portions of the relative stereochemistry of intricarene were readily assigned by NOE NMR spectral methods. However, while the relative configurations about the three spirocyclic carbons in **1** (C-2, C-6, and C-12) could be assigned indirectly on the basis of the overall correlations observed in the NOESY NMR spectrum, confirmation of the entire structure of intricarene by single-crystal X-ray diffraction analysis was highly desirable. Thus, recrystallization of **1** by slow evaporation from a mixture of $\text{MeOH}/\text{CHCl}_3/\text{acetone}$ gave crystals of excellent quality that were amenable to X-ray crystallographic analysis. The X-ray crystal structure, which defines only the relative configuration, is shown in Figure 1.⁶ Hence, the overall relative stereochemistry for the six stereocenters within the highly strained ring systems in intricarene was assigned as 1*S**, 2*R**, 6*R**, 10*S**, 11*S**, and 12*S**. The assigned configurations, which were in full agreement with the correlations observed in the NOESY spectrum of **1** (Figure 2), were confirmed through interpretation of NMR coupling constant data (Table 1).

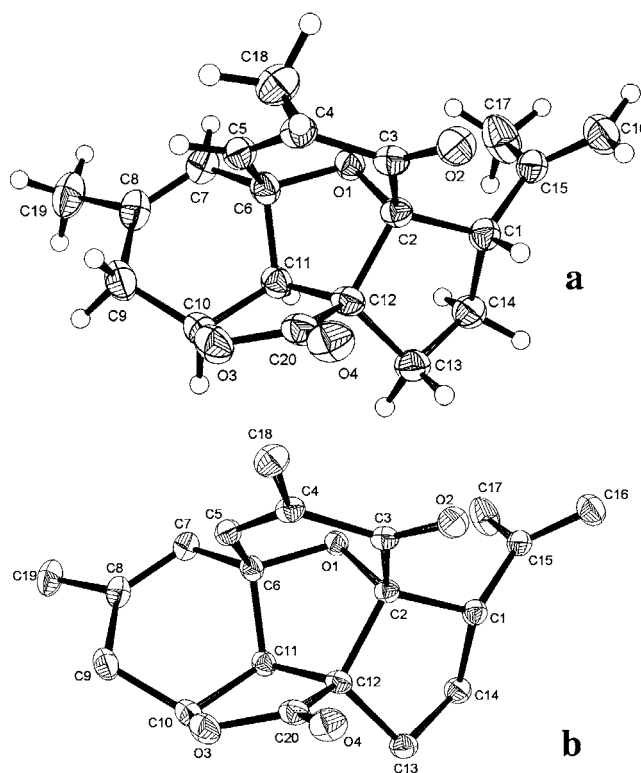


Figure 1. Computer-generated ORTEP drawings of intricarene (**1**) with atom labeling scheme. The absolute configuration shown is arbitrary. In (a) the carbon and oxygen atoms are drawn as 50% thermal ellipsoids; in (b) the hydrogen atoms are omitted for clarity, and the carbon and oxygen atoms are drawn as 30% thermal ellipsoids.

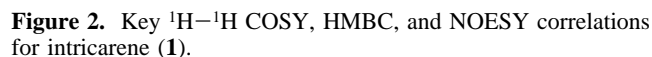
Intricarene (**1**) has an unprecedented regular diterpenoid carbon skeleton that can be formally derived from the “cembrane” skeleton found in many *Pseudopterogorgia* metabolites by sequential cyclization via [C6 \rightarrow C11] and [C2 \rightarrow C12] bond formation.⁷ The co-occurrence of **1** with various furanocembranoid lactones within the same organism suggests the biogenetic pathway outlined in Scheme 1.⁸ We suggest the semisystematic name “intricarane” to define this new carbon skeleton and a numbering scheme that preserves the C1–C20 numbering of the cembrane skeleton.

Intricarene is exceptional in several respects. While other diterpenes based on a [6–7–5] tricarbocyclic system have

(6) Crystal data for intricarene (**1**) at 298(2) K: $\text{C}_{20}\text{H}_{22}\text{O}_4$, $M_r = 326.38$, monoclinic, space group $P2_1$, $a = 7.1134(4)$, $b = 12.6053(6)$, $c = 9.4510(5)$ Å, $\beta = 105.5470(2)^\circ$, $V = 816.43(6)$ Å³, $Z = 2$, $\rho_{\text{calc}} = 1.328 \text{ Mg m}^{-3}$, $F_{000} = 348$, $\lambda(\text{Mo K}\alpha) = 0.71073$ Å, $\mu = 0.092 \text{ mm}^{-1}$. Data collection and reduction: crystal size, $0.50 \times 0.25 \times 0.25 \text{ mm}^3$, θ range, $2.24\text{--}27.16^\circ$, 5909 reflections collected, 1882 independent reflections ($R_{\text{int}} = 0.0291$), final R indices ($I > 2\sigma(I)$); $R_1 = 0.0322$, $wR_2 = 0$ for 220 variable parameters, $\text{GOF} = 1.030$. CCDC 265687 (**1**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB21EZ, UK; fax: (+44)1223-336-033; e-mail: deposit@ccdc.cam.ac.uk).

(7) (a) Wright, A. E.; Burres, N. S.; Schulte, G. K. *Tetrahedron Lett.* **1989**, 30, 3491–3494. (b) Rodríguez, A. D.; Shi, J.-G.; Huang, S. D. *J. Nat. Prod.* **1999**, 62, 1228–1237.

(8) Complete details of the isolation, purification, structure elucidation, and biological activities of all of the new furanocembranoides coisolated during the present study will soon be reported.



Compound **1** was tested for its inhibitory activity toward the growth of *Mycobacterium tuberculosis* H₃₇Rv (ATCC 27294) in the microplate Alamar Blue assay.¹⁰ The highest percentage of inhibition attained (15%) was detected at a concentration of 128 μ g/mL, thus suggesting a lack of appreciable toxicity against this bacterium strain. Unfortunately, the paucity of this material prevented us from conducting further biological screenings.

Acknowledgment. We thank Dr. Juan A. Sánchez and the staff of the Ministerio del Medio Ambiente (Bogotá, Colombia) for assistance during the collection and taxonomic identification of *P. kallos*. Dr. Scott G. Franzblau from The Institute for Tuberculosis Research (University of Illinois at Chicago) provided the in vitro antimycobacterial data for **1**. The high-resolution EI mass spectral determination was provided by the Mass Spectroscopy Laboratory of the University of Illinois at Urbana–Champaign. We gratefully acknowledge the NIH-MBRS SCORE/RISE Programs of the University of Puerto Rico for partial financial support of this work.

Supporting Information Available: Detailed description of the experimental procedures, copies of the NMR (^1H and ^{13}C), and HREI-MS spectra of intricarene (**1**), a partial mechanistic interpretation of the mass-spectral fragmentation data, and tables of crystallographic data for **1** (crystal data and structure refinement, atomic coordinates, bond lengths and angles, anisotropic displacement parameters, and hydrogen coordinates). This material is available free of charge via the Internet at <http://pubs.acs.org>.

OL0505961

- (9) (a) Dunlop, R. W.; Ghisalberti, E. L.; Jefferies, P. R.; Skelton, B. W.; White, A. H. *Aust. J. Chem.* **1989**, *42*, 315–319. (b) Brady, S. F.; Bondi, S. M.; Clardy, J. *J. Am. Chem. Soc.* **2001**, *123*, 9900–9901.
- (10) Cantrell, C. L.; Lu, T.; Fronczek, F. R.; Fisher, N. H.; Adams, L. B.; Franzblau, S. G. *J. Nat. Prod.* **1996**, *59*, 1131–1136.